

UNCLASSIFIED

AD **409 936**

DEFENSE DOCUMENTATION CENTER

FOR

SCIENTIFIC AND TECHNICAL INFORMATION

CAMERON STATION, ALEXANDRIA, VIRGINIA



UNCLASSIFIED

NOTICE: When government or other drawings, specifications or other data are used for any purpose other than in connection with a definitely related government procurement operation, the U. S. Government thereby incurs no responsibility, nor any obligation whatsoever; and the fact that the Government may have formulated, furnished, or in any way supplied the said drawings, specifications, or other data is not to be regarded by implication or otherwise as in any manner licensing the holder or any other person or corporation, or conveying any rights or permission to manufacture, use or sell any patented invention that may in any way be related thereto.

409 936

63-4-2

FINAL **REPORT ON**

CONTRACT NO DA 92-557-FEC-35805

EXCLUSIVE DATES February 1, 1962 TO January 31, 1963

SUBJECT OF INVESTIGATION

The explanation of pathogenesis of Japanese encephalitis (JE) virus and the establishment of an attenuated strain of JE virus completely a virulent to man

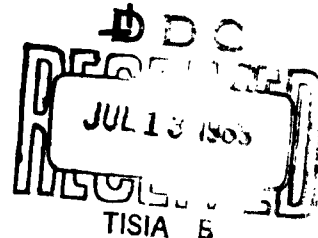
Part III

Studies on Japanese encephalitis virus infection within cultivated animal cells by the application of fluorescent antibody technique

RESPONSIBLE INVESTIGATOR

Akira Oya, M.D.

Chief of the 4th Virus Division,
Department of Virology & Rickettsiology,
National Institute of Health, Tokyo, Japan.



U.S. Army Research & Development Group (9852) (Far East)

Office of the Chief of Research and Development

United States Army

APO 343

CATALOGED BY DDC
AS AD No.

D-I-S-T-R-I-B-U-T-I-O-N

The distribution of this report as made by USA R&D Gp (FE)
is as follows:

Army Research Office, 3045 Columbia Pike, Arlington 4, (4)
Virginia. ATTN: Chief Research Programs Office

Army Attache, American Embassy, Tokyo, Japan (1)

U. S. Army Medical R & D Command (4)

ASTIA (10) ✓

~~Office of Primary Scientific Liaison~~ ()

~~Offices of Scientific Cognizance~~ ()

THE EXPLANATION OF PATHOGENESIS OF JAPANESE ENCEPHALITIS
(JE) VIRUS AND THE ESTABLISHMENT OF AN ATTENUATED STRAIN
OF
JE VIRUS COMPLETELY A VIRULENT TO MAN

PART III

STUDIES ON JAPANESE ENCEPHALITIS VIRUS INFECTION
WITHIN
CULTIVATED ANIMAL CELLS
BY THE APPLICATION OF FLUORESCENT ANTIBODY TECHNIQUE

Akira Oya, M.D.

Chief of 4th Virus Division,
Department of Virology & Rickettsiology,
National Institute of Health, Tokyo, Japan.

C O N T E N T S

| | Page |
|---------------------------------------|------|
| a. Purpose of Study | 1 |
| b. Materials and Methods | 2 |
| c. Experimental Results | 3 |
| 1. Infection of HK cells | 3 |
| 2. Infection of PS cells | 4 |
| 3. Infection of HeLa cells | 5 |
| d. Discussion | 6 |
| e. References | 7 |
| f. Abstract of the Final Report | 8 |
| Explanation of Plates | 9 |

The explanation of pathogenesis of Japanese encephalitis (JE) virus and the establishment of an attenuated strain of JE virus completely a virulent to man

Part III

Studies on Japanese encephalitis virus infection within cultivated animal cells by the application of fluorescent antibody technique

a. Purpose of Study

Japanese encephalitis virus infection of mammals are known to result in a variety of sequences. A large part of infected animals does not show any sign of illness to severe involvement of the central nervous system. Originally, our study was attempted to know some fundamental rules of JE infection sequences occurring in the mammals.

Previous report, part II of the investigation under the same title, described the evidences which suggested the following infection process in the peripherally infected adult mice. Peripheral infection induces firstly the viral growth some where outside of brain tissue. Viremia ensues from the primary viral growth and provides a cause of viral invasion to brain tissue. Those two steps, primary viral growth and viremia seem to be essential form of JE infection of the adult mice. On the other hand, the reason why encephalitis occur in some mice and the other mice remain in apparently healthy lookings is still quite obscure.

The next aim of our investigation was thought to clarify kinds of tissues or cells involved in the viral growth not only in the encephalitic phase but in the visceral stage. Recent development of fluorescent antibody technique seemed to provide a reasonable tool to approach to the present aim. However, no work has ever been done with this method for JE virus study.

Therefore, it was considered to be wise to begin with the simple system as possible, i.e. tissue culture system before applying it to more complicated tissues. Present paper deals with the results so far obtained in the cultivated cells infected with JE virus.

b. Materials and Methods

Virus strain: Nakayama-NIH strain of JE virus of repeated mouse brain passages more than hundred times was adapted to hamster kidney (HK) cells by approximately 10 passages was used.

Cultivated cells: Three kinds of cells, i.e. secondary transferred HK cells, continuous pig kidney (PS) cells and HeLa cells were employed. Those cells were reported to be highly susceptible to JE virus demonstrating cell degeneration (Kissling, 1957; Inoue et al, 1962; Buckley, 1959). Cell suspensions of 1×10^5 cells per ml for HK and PS and 2×10^5 cells per ml for HeLa were seeded onto cover slips (12 x 32mm) kept in petri dishes. Dishes were incubated at 37°C for 2 days under the continuous supply of 5% CO₂ gas. Then, cells on cover slips were washed twice with phosphate buffered saline (PBS) and transferred to small square tubes with 0.9 ml of maintenance medium.

Growth media for cell culture: For HK and PS cells, 10% calf serum plus 0.5 La-Hanks BSS and for HeLa, 20% calf serum plus 0.1% yeast extract, 0.5% La-Earle BSS were used.

Maintenance medium: Four per cent calf serum free of JE antibody plus 0.5% La-Hanks BSS was used through all the experiment.

Inoculation and harvest: One tenth ml of infected HK fluid bearing JE virus titer of 10^7 TCID₅₀ per ml was inoculated into each 0.9 ml of maintenance medium of the experimental cell tubes. The input multiplicity was calculated to be between 5 to 10 so as to obtain

such condition that large number of cells were infected simultaneously. At suitable hours of infection described in the text, 2 tubes were randomly selected from the infected group and uninfected controls, and cover slips were taken out. They were washed with saline and Hanks BSS, then fixed in dried acetone at room temperature. Fixed preparations were dried on filter paper and kept at -20°C until antibody treatment.

Preparation of anti-JE virus antiserum: Five day chick embryo suspension infected with JE virus JATH 160 strain was used for immunization of rabbits several times. Thus immunized rabbit serum was found to contain HI antibody 1:400 against 8 units of Nakayama HA antigen.

Preparation of conjugated anti-rabbit globulin
Goat serum: Gamma globulin was purified from normal rabbit serum, mixed with Freund's adjuvant and injected into goat subcutaneously. Thus, obtained goat serum was conjugated with fluorescein isocyanate.

Staining and observation of the specimen: Anti-JE virus antiserum was diluted to 1:20 and mounted on the fixed cells on cover slips for 30 min at room temperature. Cover slips were then washed with saline and mounted with conjugated serum for another 30 min. After washing off serum, cells on cover slips were observed by Reichert's fluorescent microscope.

C. Experimental Results

1. Infection of HK cells.

Cells of uninfected controls were stained by fluorescein faint and diffuse both in cytoplasm and nucleus (Pl. 1 and 2). It was obvious at our present experiment that some non-viral or non-specific staining

was unavoidable. However, it should be emphasized that there was no difference in the illumination of cytoplasm and nucleus. At 2, 6 hours of infection, no difference could be observed with cells between infected and uninfected groups. Remarkable difference was at first noted at 10 hrs of infection. As clearly shown in Plates 3 and 4 marked illuminated boundary was observed around the nucleus in some cells. In contrast to the above boundary, the nucleus appeared as a dark spot. At 15 hrs., bright area expanded in cytoplasm from perinuclear site. Illuminated area seemed diffuse and consisted of many fine granules. Number of illuminated cells became more increased. Whole part of cytoplasm became stained in most of cells after 24 hours of infection (Pl. 5). Granules in the illuminated area appeared more coarse than before. It should be pointed out that the nuclei were still remained unstained. At this period, cells began to indicate degenerative changes, i.e. projection, shrinkage and leakage of cytoplasm. Coincidentally with cytopathic changes, viral infective titer in the culture fluid increased abruptly.

2. Infection of PS cells.

In the earlier stage of infection, infected cells were stained by fluorescent faint and diffuse (Pl. 7) and no difference could be observed between infected and control cells (Pl. 6). It was also pointed out that little difference was noted in the brightness of nucleus and cytoplasm of the cell. Appearance of the infected cells differed remarkably at first 10 hours after the virus inoculation (Pl. 8). Cytoplasms of some part of infected cells began to emit remarkable fluorescent light. Loci of the illuminated areas were restricted to the perinuclear area. However, it did not appear as a sharp boundary observed in HK cells.

Number of the illuminated cells increased at 15 hours (Pl. 9). At this time, most of cells seemed to contain bright area. Within a single cell, bright area consisted of fine granules occupied the whole cytoplasm. The fact should be emphasized that nucleus remain dark in quite a contrast to the cytoplasm. No degenerative process could be observed by this time. Cytophathy was noticed from 24 hours of infection. At this stage, coarse particles were marked in the light scattering area in cytoplasm (Pl. 10). Though some illuminated material could be seen in nucleus, it would be explained by cytoplasmic materials covering the top of the nucleus.

3. Infection of HeLa cells.

Pictures obtained with HeLa cells were essentially the same with those observed with PS cells. Only the difference we noticed between above two cell systems was the fact that the infection proceeded slower in HeLa than in PS. First recognition of specific viral antigen in HeLa cell was at 15 hours after the inoculation as seen in Plate 12. Control cells shown in Plate 11 will give the idea that non specific staining seems a little stronger in HeLa. However it should be kept in mind that exposing time with control specimen was 2 to 3 times longer than that with other infected pictures. Most of cells became illuminated at 24 hours and lasted without sign of degeneration for more than 24 hours (Pl. 13,14). Antigen accumulation was restricted to cytoplasmic area and nucleus seemed completely unstainable. Degeneration of cells appeared 72 hours of infection. Illuminated area seemed to localize within the cytoplasm and coarse particles became remarkable at this stage (Pl. 15).

d. Discussion

The purpose of the present investigation is to know the applicability of fluorescent antibody technique to JE virus-cell system and to know the general rule of viral antigen accumulation inside the cell. It has been known with tissue culture of JE virus that there are two types of cells susceptible to JE virus. The one shows degenerative changes by the infection and another appears completely unaffected, though both types of cells render JE virus to grow inside the cells. HK and PS cells were known to belong to the former cytopathogenic type. However, the opinion has not been in accord with HeLa cells. Scherer and Syverton (1954) and McCollum and Foley (1957) reported inconsistent cytopathic effect with HeLa cells infected with JE virus. On the contrary, Buckeley (1959) reported consistent cytopathic effect with this system. The latter author seemed to attribute this difference to the difference of the medium in which cells were propagated. However, HeLa cells which were used in the present experiment was found highly degenerative after the JE virus infection. Thus, cytopathic changes enabled us viral titration on this cell line. Therefore, three kinds of cells used in the present experiment showed eventually cytopathic changes.

On the site of antigen production of JE virus inside the cell. Prince (1960) suggested some participation of cell nucleus observing specific fluorescent illumination inside the nucleus at earlier stage of infection to HK cells. On the contrary, Noyes (1955) described no evidence of antigen localized in the nucleus of mouse cortex or human carcinoma cells infected with West Nile virus which was antigenically related to JE virus. In the present report, JE antigen was found strictly in cytoplasm at every stage of

infection and no evidence was obtained whatsoever to suggest the localization of JE antigen within cell nucleus similarly to Noyes's report. Nevertheless, it would not be correct to conclude that cell nucleus has no relation to antigen production of JE virus. Provided that our observation of the illuminated perinuclear boundary is specific for viral antigen, it seems still probable that cell nucleus may take some part for the initiation of viral antigen reproduction. Although such marked boundary could not be found in cases of PS and HeLa infections, it should be emphasized that no antigen producing site could be observed completely independent to the nucleus in the earlier stage of infection.

As a whole, the pictures elucidated in the present report seems quite similar to those reported in West Nile, Dengue(Buckeley, 1961) viruses suggesting the existence of common rule in the reproduction of arbor viruses. It was also confirmed that fluorescent antibody technique could furnish a convenient tool for the investigation of pathogenesis of JE virus.

e. References.

- Buckeley, S.M., 1959. Propagation, cytopathogenicity, and hemagglutination-hemadsorption of some arthropod-borne viruses in tissue culture. *Annals New York Acad. Sci.*, 81, 172 - 187.
- Buckeley, S.M., 1961. Serial propagation of types 1, 2, 3 and 4 dengue virus in HeLa cells with concomitant cytopathic effect. *Nature*, 192, 778 - 779.
- Inoue Kanda, Y. and Ogura, R. 1962. Studies on Japanese B encephalitis virus. III. Propagation and assay of Japanese B encephalitis virus in a stable line of porcine kidney cells. *Virology*, 16, 205 - 207.

- Kissling, R.E. 1957. Growth of several arthropod-borne viruses in tissue culture. Proc. Soc. Exp. Biol. & Med., 96, 290 - 294.
- McCollum, R.W. and Foley, J.F. 1957. Japanese B encephalitis virus in tissue culture. Proc. Soc. Exp. Biol. & Med., 94, 556 - 550.
- Noyes, W.F., 1955. Visualization of Egypt 101 virus in the mouse's brain and in cultured human carcinoma cells by means of fluorescent antibody. J. Exp. Med., 102, 243 - 248.
- Prince, A.M. 1960. personal communication.
- Scherer, W.F. and Syverton, J.T. 1954. The viral range in vitro of a malignant human epithelial cell (Strain HeLa, Gay). II. Studies with encephalitis viruses of the eastern, western, West Nile, St. Louis and Japanese B types. Am. J. Path., 30, 1075 - 1084.

f. Abstract of the Final Report.

Process of Japanese encephalitis virus infection in cultivated animal cells, i.e. hamster kidney, porcine kidney and HeLa cells was followed by fluorescent antibody technique. The infection was adjusted to obtain multiplicities 5 - 10 for each system. Viral antigen was firstly detected in perinuclear cytoplasm at 10 - 15 hrs of infection, then numbers of the antigen producing cells increased and antigen producing area expanded within cytoplasm. No evidence was obtained that antigen localized in nucleus. Infected cells were eventually degenerated in all cell systems. Applicability of fluorescent antibody technique to the investigation of pathogenesis was considered hopeful.

Explanation of Plates

- Plate 1. Uninfected control of HK cells. 200 X.
Exposure: 20".
- Plate 2. Uninfected control of HK cells. 500 X.
Exposure: 30".
- Plate 3. HK cells at 10 hrs after the inoculation.
200 X. Exposure: 30".
- Plate 4. HK cells at 10 hrs after the inoculation.
500 X. Exposure: 30".
- Plate 5. HK cells at 24 hrs after the inoculation.
200 X. Exposure: 10".
- Plate 6. Uninfected PS cells. 200 X. Exposure: 80".
- Plate 7. PS cells at 6 hrs after the inoculation.
200 X. Exposure: 40".
- Plate 8. PS cells at 10 hrs after the inoculation.
200 X. Exposure: 45".
- Plate 9. PS cells at 15 hrs after the inoculation.
200 X. Exposure: 45".
- Plate 10. PS cells at 27 hrs after the inoculation.
200 X. Exposure: 30".
- Plate 11. Uninfected HeLa cells. 200 X. Exposure:
120"
- Plate 12. HeLa cells at 15 hrs after the inoculation.
200 X. Exposure: 60".
- Plate 13. HeLa cells at 24 hrs after the inoculation.
200 X. Exposure: 60".
- Plate 14. HeLa cells at 48 hrs after the inoculation.
200 X. Exposure: 30".

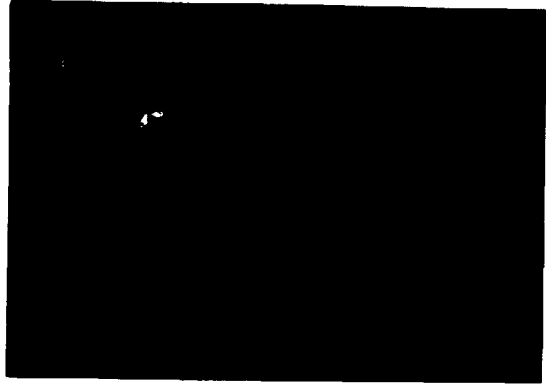


Plate 1



Plate 2



Plate 3



Plate 4

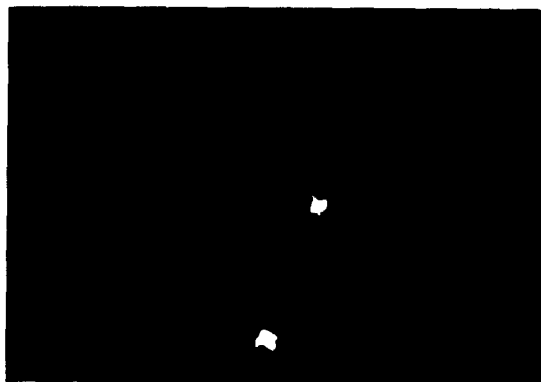


Plate 5



Plate 6



Plate 7



Plate 8



Plate 9

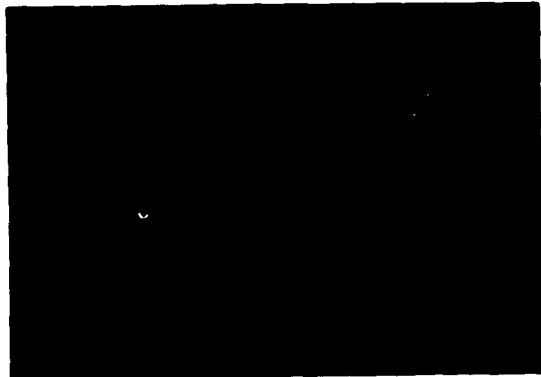


Plate 10



Plate 11



Plate 12



Plate 13

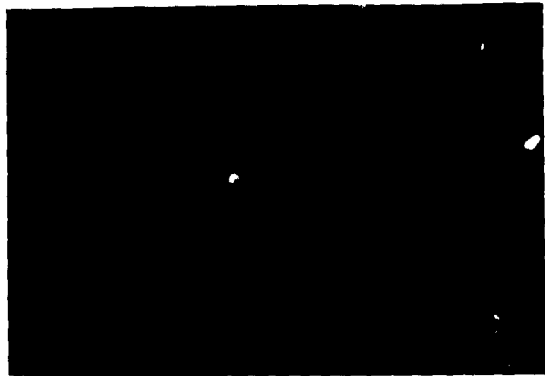


Plate 14



Plate 15

AD Div 16/2

National Institute of Health, Tokyo (Japan)
STUDIES ON THE PATHOGENESIS OF JAPANESE B
ENCEPHALITIS VIRUS AND ESTABLISHMENT OF A
JAPANESE B ENCEPHALITIS VIRUS STRAIN COMPLETELY
ADAPTED TO MAN, by Akira Oya.
Final Report No. 1, 1 Feb 62 to 31 Jan 63. 9 p.
incl illus, tables, 8 refs.
(Contract DA 92-557-FRC-35805)Unclassified report

Process of Japanese B encephalitis virus in-
fection in cultivated animal cells. i.e. hamster
kidney, porcine kidney and HeLa cells was fol-
lowed by fluorescent antibody technique. The
infection was adjusted to obtain multiplicities
5-10 for each system. Viral antigen was initial-
ly detected in perinuclear cytoplasm at 10 - 15
hrs of infection, then numbers of the antigen
producing cells increased and antigen producing
area expanded within cytoplasm.

AD Div 16/2

National Institute of Health, Tokyo (Japan)
STUDIES ON THE PATHOGENESIS OF JAPANESE B
ENCEPHALITIS VIRUS AND ESTABLISHMENT OF A
JAPANESE B ENCEPHALITIS VIRUS STRAIN COMPLETELY
ADAPTED TO MAN, by Akira Oya.
Final Report No. 1, 1 Feb 62 to 31 Jan 63. 9 p.
incl illus, tables, 8 refs.
(Contract DA 92-557-FRC-35805)Unclassified report

Process of Japanese B encephalitis virus in-
fection in cultivated animal cells. i.e. hamster
kidney, porcine kidney and HeLa cells was fol-
lowed by fluorescent antibody technique. The
infection was adjusted to obtain multiplicities
5-10 for each system. Viral antigen was initial-
ly detected in perinuclear cytoplasm at 10 - 15
hrs of infection, then numbers of the antigen
producing cells increased and antigen producing
area expanded within cytoplasm.

UNCLASSIFIED

1. Encephalitis Viruses
- I. Title: None
- II. Oya, Akira
- III. U.S. Army Research and Development Gp (FBI), OCRD, DA, Wash, D. C.
- IV. Contract DA 92-557-FRC-35805

Armed Services
Technical Information
Agency

UNCLASSIFIED

AD

Div 16/2

National Institute of Health, Tokyo (Japan)
STUDIES ON THE PATHOGENESIS OF JAPANESE B
ENCEPHALITIS VIRUS AND ESTABLISHMENT OF A
JAPANESE B ENCEPHALITIS VIRUS STRAIN COMPLETELY
ADAPTED TO MAN, by Akira Oya.
Final Report No. 1, 1 Feb 62 to 31 Jan 63. 9 p.
incl illus, tables, 8 refs.
(Contract DA 92-557-FRC-35805)Unclassified report

Process of Japanese B encephalitis virus in-
fection in cultivated animal cells. i.e. hamster
kidney, porcine kidney and HeLa cells was fol-
lowed by fluorescent antibody technique. The
infection was adjusted to obtain multiplicities
5-10 for each system. Viral antigen was initial-
ly detected in perinuclear cytoplasm at 10 - 15
hrs of infection, then numbers of the antigen
producing cells increased and antigen producing
area expanded within cytoplasm.

AD

Div 16/2

National Institute of Health, Tokyo (Japan)
STUDIES ON THE PATHOGENESIS OF JAPANESE B
ENCEPHALITIS VIRUS AND ESTABLISHMENT OF A
JAPANESE B ENCEPHALITIS VIRUS STRAIN COMPLETELY
ADAPTED TO MAN, by Akira Oya.
Final Report No. 1, 1 Feb 62 to 31 Jan 63. 9 p.
incl illus, tables, 8 refs.
(Contract DA 92-557-FRC-35805)Unclassified report

Process of Japanese B encephalitis virus in-
fection in cultivated animal cells. i.e. hamster
kidney, porcine kidney and HeLa cells was fol-
lowed by fluorescent antibody technique. The
infection was adjusted to obtain multiplicities
5-10 for each system. Viral antigen was initial-
ly detected in perinuclear cytoplasm at 10 - 15
hrs of infection, then numbers of the antigen
producing cells increased and antigen producing
area expanded within cytoplasm.

UNCLASSIFIED

1. Encephalitis Viruses
- I. Title: None
- II. Oya, Akira
- III. U.S. Army Research and Development Gp (FBI), OCRD, DA, Wash, D. C.
- IV. Contract DA 92-557-FRC-35805

Armed Services
Technical Information
Agency

UNCLASSIFIED

UNCLASSIFIED

1. Encephalitis Viruses
- I. Title: None
- II. Oya, Akira
- III. U.S. Army Research and Development Gp (FBI), OCRD, DA, Wash, D. C.
- IV. Contract DA 92-557-FRC-35805

Armed Services
Technical Information
Agency

UNCLASSIFIED

UNCLASSIFIED
DESCRIPTORS
Virus
Encephalitis Viruses
Antigens
Mice
Japan
Viability
Separation
Antibodies
Infections
Mutations

No evidence was obtained that antigen localized in nucleus. Infected cells were eventually degenerated in all cell systems. Applicability of fluorescent antibody technique to the investigation of pathogenesis was considered hopeful. (Author)

UNCLASSIFIED
DESCRIPTORS
Virus
Encephalitis Viruses
Antigens
Mice
Japan
Viability
Separation
Antibodies
Infections
Mutations

No evidence was obtained that antigen localized in nucleus. Infected cells were eventually degenerated in all cell systems. Applicability of fluorescent antibody technique to the investigation of pathogenesis was considered hopeful. (Author)

UNCLASSIFIED
DESCRIPTORS
Virus
Encephalitis Viruses
Antigens
Mice
Japan
Viability
Separation
Antibodies
Infections
Mutations

No evidence was obtained that antigen localized in nucleus. Infected cells were eventually degenerated in all cell systems. Applicability of fluorescent antibody technique to the investigation of pathogenesis was considered hopeful. (Author)

UNCLASSIFIED
DESCRIPTORS
Virus
Encephalitis Viruses
Antigens
Mice
Japan
Viability
Separation
Antibodies
Infections
Mutations

No evidence was obtained that antigen localized in nucleus. Infected cells were eventually degenerated in all cell systems. Applicability of fluorescent antibody technique to the investigation of pathogenesis was considered hopeful. (Author)

UNCLASSIFIED

UNCLASSIFIED